**Antimicrobial potential of leaves of Psidium guajava.**

Chandekar C.J\*; M.J.Madhugiri.

Department of Microbiology and Biotechnology,SSES Amt’s Science College,congress nagar, Nagpur-440 012,Maharashtra,India.

\*Corresponding author’s email:chandekarc@yahoo.com.

**ABSTRACT**

The aim of the study was focused on the antibacterial potential of dried leaves solvents extracts of five different extracts (acetone, chloroform, methanol, petroleum ether and water) of *Psidium guajava*(family Myrtaceae) were tested against *Bacillus cereus*, *Bacillus subtilis*,*Bacillus megaterium,* *Staphylococcus aureus*, *Escherichia coli*, *Proteus vulgaris* and *Pseudomonas aeruginosa* by well diffusion method.Acetone, methanol and petroleum ether extracts showed significant antibacterial activity.All the organisms are found susceptible except *Bacillus cereus*. Chloroform extract is active against only *Bacillus subtilis.* The fresh leaves juice was found to be active against all organisms except *Pseudomonas aerugenosa* and the aqueous extract found to be active against *Proteus vulgaris* and *Staphylococcus aureus..*Results were compared with standard antibiotics Amoxicillin-Am30, Ciprofloxacin-Cf30, Cotrimaxazole-Co25,Gentamicin-G50 and Tetracycline-T30 .

**KEYWORDS:** *Psidium guajava,* solvent extracts, antibacterial sensitivity test.

**INTRODUCTION**

In developing countries,the frequency of life threatening infections were caused by pathogenic microorganisms has led to increased worldwide and in becoming an important cause of morbidity and mortality in immunocompromised patients ( Al-Bari et al.,2006 ).The historical point, plants have been used as an important source of natural products for human health.All over the world, the antimicrobial properties of plants have been investigated by a number of studies and many of them have been used as therapeutic alternatives because of their antimicrobial properties( Adriana et al.,2007) and they contain secondary metabolites such as alkaloids,phenolic compounds,etc.The practice of complementary and alternative medicine is now on the increase in developing countries in response to World Health Organization directives culminating in several pre-clinical and clinical studies that have provided the scientific basis for the efficacy of many plants used in folk medicine to treat infections( Vijaya and Ananthan, 1997; and Dilhuydy, 2003).It is therefore very necessary that the search for newer antibiotic sources be a continuous process.Plants are the cheapest and safer alternative sources of antimicrobials(Pretorius and Watt, 2001; Doughari et al.,2007). According to World Health Organization (Santos et al.,1995) medicinal Plants would be the best source to obtain a variety of drugs.About 80% of individuals from developed countries use traditional medicine, which has compounds derived from medicinal plants. Therefore, such plants should be investigated to better understand their properties, safety and efficacy(Ellof,1998). In India thousands of species are known to have medicinal values and the use of different parts of several medicinal plants to cure specific ailments has been in vogue since ancient times( Parekh et al.,2005) Medicinal plants are valuable natural resources and regarded as potentially safe drugs and have been tested for biological, antimicrobial and hypoglycemic activity also play an important role in modern medicine(Hassawi and Kharma,2006; Bhat et al.,2009) . It is well known that even the most synthetic drugs have their origin from plant products( Sofowara,1982 ) . The screening of plant extracts and their products for antimicrobial activity has shown that higher plants represent a potential source of novel antibiotics prototypes(Afolayan,2003) The selection of crude plant extracts for screening programs is potentially more successful in initial steps than the pure compounds( Kasamato et al.1995). Such screening of various plant extracts has been previously studied by many workers(Erdogrul,2002; Parekh and Chanda ,2007). Eventhough hundreds of plant species have been tested for antimicrobial properties,the vast majority of them have not yet been evaluated.

*Psidium guajava* L,commonly known as guava ,of the family Myrtaceae,is a native plant of India.Different parts of the plant are used in the indigenous system of medicine for the treatment of various human ailments such as wounds,ulcers,bowels and cholera(Begum et al.,2002).Pharmacological investigations indicated that its bark, fruit, and leaves possess antibacterial, hypoglycemic, anti-inflammatory, analgesic, antipyretic, spasmolytic, and CNS depressant activities(Begum et al.,2002).In Mexico,*P.guajava* leaves are extensively used to stop diarrhea,and the quercetin and its glycosides were its active compounds.The leaves of P.guajava contain an essential oil rich in cineol, tannins, triterpenes and flavonoids(Olajide et al.,1999).

In vitro studies on plants used in traditional medicine have been carried out in the field of microbiology, especially on pathogenic bacterial growth; and some studies were about the antimicrobial activity of *Psidium guajava* (Gnan and Demello, 1999; Jairaj et al.,1999; Nascimento et al.,2000; Ahmad and Beg, 2001; Abdelrahim et al.,2002; Holetz et al.,2002; Voravuthikunchai et al.,2004; Qadan et al.,2005).

The present study was undertaken to investigate the in vitro antibacterial activity of water, acetone, chloroform, methanol and petroleum ether extracts from leaves of *Psidium guajava*.

**MATERIALS AND METHODS**

**Selection of medicinal plant for this study**

**Identification and Preservation of Plant materials:**

Fresh plant leaves of *Psidium guajava* were collected from the Nagpur area and identified.Plant leaves were washed with 70% alcohol and then rinsed with sterilized distilled water, air dried and stored in airtight bottles at 4oC for further use.

**Preparation of crude extract**

Homogenized mass of leaves was squeezed in 400 mesh nylon cloth (pore size 37 micron) to obtain crude extract. Crude extract was prepared fresh and used before 2 hr. Cold extracts were prepared using individual fresh plant leaves.

**Crude Extraction**

**Aqueous extraction**

Ten gram of dried powder was extracted in 100 ml distilled water for 6 hr at slow heat. Every 2 hr, it was filtered through 8 layers of muslin cloth and centrifuged at 5000g for 15 min. The supernatant was collected. This process was repeated twice and after 6 hr, the supernatant wasconcentrated to make the final volume one-fourth of the original volume (Shahidi Bonjar, 2004). It was then autoclaved at 121 ºC and 15 lbs pressure and then stored at 4ºC.

**Solvent extraction**

Ten gram of dried powder was extracted with 100 ml of each solvent (acetone, chloroform, methanol and petroleum ether)and flasks were kept on a rotary shaker at 190-220 rpm for 24 hr. Thereafter, it was filtered through 8 layers of muslin cloth and centrifuged at 5000 g for 15 min. The supernatant was collected and the solvent was evaporated to make the final volume one-fourth of the original volume (Shahidi Bonjar, 2004). It was stored at 4oC in airtight bottles for further studies.

**Bacterial cultures**

The microbial strains are identified strains and were obtained from the National Chemical Laboratory (NCL),Pune, India. The studied bacterial strains were *Bacillus cereus* NCIM2155, *Bacillus subtilis* NCIM2063,*Bacillus megaterium* NCIM2087, *Escherichia coli* NCIM2931, *Proteus vulgaris* NCIM2857 and *Pseudomonas aeruginosa*NCIM5029. *Staphylococcus aureus* MTCC96 this strain was procured from Institute of Microbial Technology(IMTECH),Chandigarh,India.They were sub-cultured on nutrient agar for every 15 days and maintained on nutrient agar slants at 40C.

**Media:**

Hi -Sensitivity test broth (M 486) and Hi-sensitivity test agar (M 485) were procured from Hi-media Mumbai,India. The media were prepared according to the instructions given.

**Screening for the antimicrobial potential of the plant leaves extracts**

The antimicrobial activity of different solvent extracts was evaluated by agar well diffusion(Perez ,et.al., 1990;and Parekh,et al.,2007) using Hi-sensitivity test agar (M 485).

**Preparation of inoculum** – A loopful of culture was inoculated from the stock slant culture in 5 ml of Hi-sensitivity test broth and broth was incubated at 35±0.5**0**C in incubator for 18-20 hr.

After incubation a loopful of actively growing culture was inoculated into 10 ml of Hi-sensitivity broth. Broth was incubated at 35±0.5**0**C for 6-8 hr. This culture was used for the inoculation of Hi-sensitivity test agar plates.

**Preparation of Hi-sensitivity test agar medium**

Hi-sensitivity test agar medium was prepared as per instructions of manufacturer. Required amount of agar medium was melted and 25 ml of molten medium was distributed in test tubes (25x150 mm). Medium was autoclaved at 15 lb. for 20 min. After autoclaving, medium was maintained at 45-50**0**C in constant temperature water bath.

**Inoculation of medium with test organism**

0.5 ml of 6-8 hours old test organism is transferred to petridish of 100 mm size(Sterilized in oven at 180**0**C for 1 hr.) using sterile micropipette. Hi-sensitivity test agar medium maintained at 45-50**0**C was poured and mixed properly to ensure uniform distribution of organisms with medium. Seeded plates are allowed to set at room temperature.

**Preparation of agar well for fresh leaves juice**

10 mm borer was used to prepare wells in agar. Four wells per plate at four equidistant corners were made.A 100 µl crude extract(fresh leaves juice) was transferred by micropipette per well. Plates were immediately kept at 40C in refrigerator for 1 hr.for the diffusion of extract and then shifted to 35±0.50C in incubator. Zone of inhibition was measured by zone scale after 24 hr. of incubation.

**Preparation of agar wells for different solvent extracts**

5 mm borer was used to prepare wells in agar. Four wells per plate at four equidistant corners were made.A 50 ul solvent extract was transferred by micropipette per well. Plates were immediately kept at 40C in refrigerator for 1 hr for the diffusion of extract. and them shifted to 350C+0.50C in incubator. Zone of inhibition was measured after 24 hr. of incubation.For each bacterial strain, controls were maintained in which pure solvents were used instead of the extract.The control zones were sustracted from the test zones and the resulting zone diameter is obtained.

**RESULTS**

All the microorganisms responded differently to the various plant extracts and standard antibiotics.All the plant extracts and antibiotics tested showed some antimicrobial activity(Table-1).The fresh leaves juice was found to be active against all organisms except *Pseudomonas aerugenosa.*When we compared the activity of aqueous extract with fresh leaves juice, the fresh leaves juice is more active.The aqueous extract found to be active against *Proteus vulgaris* and *Staphylococcus aureus.*

Acetone ,methanol and petroleum ether extracts are active against almost all microorganisms except *Bacillus cereus.*Chloroform extract is active against only *Bacillus subtilis(Fig-3).*All the organisms are susceptible to Ciprofloxacin- Cf30 , Gentamicin-G50 ,and Tetracycline-T30.*Proteus vulgaris* is found to be resistant to Amoxycilin Am30 *Pseudomonas aerugenosa* and *Bacillus cereus* found to be resistant to Cotrimaxozole Co25.

Table-1.Results of antimicrobial activities of fresh leaves juice and solvent extracts of *Psidium guajava* and compared with standard antibiotics.

|  |
| --- |
|  |

Sr.No. Microorganisms Zone of inhibition in mm

Leaves extracts Standard antibiotics

-------------------------------------- ----------------------------------

FJ WE AE ME CE PE Am30  Cf30 Co25 G50  T30

|  |
| --- |
|  |

1.*Escherchia coli*  20 --- 17 17 --- 16 32 29 24 17 22

2. *Proteus vulgaris* 27 11 15 15 --- 16 --- 23 31 20 24

3*.Pseudomonas aerugenosa*  --- --- 21 18 --- 18 14 36 --- 34 22

4.*Staphylococcus aureus* ND 12 15 13 --- 13 31 23 20 16 17

5. *Bacillus cereus* 20 --- --- --- --- --- 15 27 --- 23 24

6.*Bacillus subtilis* 13 --- 16 16 15 14 31 50 36 40 32

7.*Bacillus megaterium* 12 --- 11 10 --- 10 29 46 24 23 33

|  |
| --- |
|  |

Key: FJ—Fresh juice of leaves; WE—Water extract; AE—Acetone extract;ME—Methanol extract;PE—Petroleumether extract ;Am30--Amoxycilin; Cf30 –Ciprofloxacin; Co25 –Cotrimaxazole; G50  --Gentamicin; Tetracycline--T30

ND—Not determined; --- Negative.

**Antibacterial activity of different solvent extracts of leaves of *Psidium guava (PG),* zone of inhibition in millimetre (mm).**

** **

**Figure-1 Figure-2**

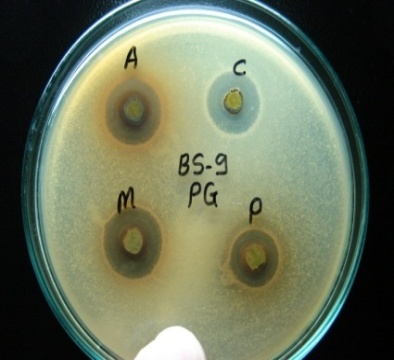
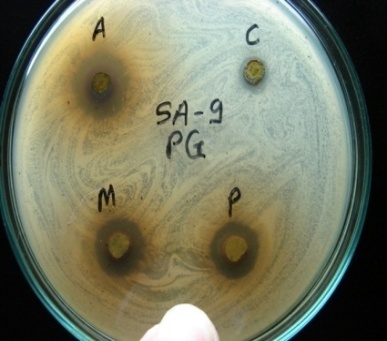
**Activity against *Proteus vulgaris* Activity against *Escherchia coli***

**Acetone extract (A)-15 mm Acetone extract (A)-17 mm**

**Chloroform extract (C) --- Chloroform extract (C) -----**

**Methanol extract (M)-15 mm Methanol extract (M)-15 mm**

**Petroleum ether extract (P)-16 mm Petroleum ether extract (P)-16 mm**

**    
 Figure-3 Figure-4**

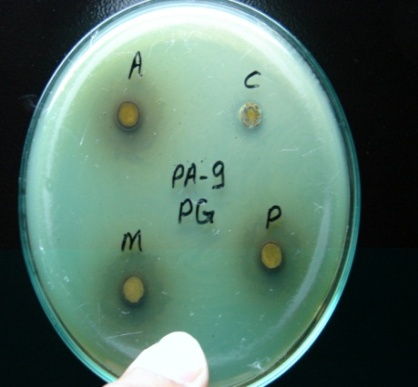
**Activity against *Bacillus subtilis* Activity against *Staphylococcus aureus***

**Acetone extract (A)-16 mm Acetone extract (A)-15 mm**

**Chloroform extract (C)-15 mm Chloroform extract (C) -----**

**Methanol extract (M)-16 mm Methanol extract (M)-13 mm**

**Petroleum ether extract (P)-14 mm Petroleum ether extract (P)-13 mm**

** **

**Figure-5 Figure-6**

**Activity against *Bacillus megaterium* Activity against *Pseudomonas aeruginosa***

**Acetone extract (A)-11 mm Acetone extract (A)-21 mm**

**Chloroform extract (C) ---- Chloroform extract (C) ----**

**Methanol extract (M)-10 mm Methanol extract (M)-18 mm**

**Petroleum ether extract (P)-10 mm Petroleum ether extract (P)-18 mm**

**DISCUSSION**

Successful prediction of botanical compounds from plant material is largely dependent on the type of solvent used in the extraction procedure.Traditional medicinal plants are used primarily water as the solvent but in our studies we found that plant extracts in organic solvent (acetone, methanol and petroleum ether) provided more consistent antimicrobial activity compared to those extracted in water. These observations can be rationalized in terms of the polarity of the compounds being extracted by each solvent and in addition to their intrinsic bioactivity.

The traditional healers use primarily water as the solvent but Suresh, et al.,2008 found in their study the plant extracts by chloroform provided less consistent antimicrobial activity compared to those extracted by water and other solvents. In our study chloroform extract is found to be active against only *Bacillus subtilis.*It has been shown that chloroform and its impurities CH2Cl2 andCH2ClBr,may react with some compounds as in the case of certain alkaloids(e.g.brucine, strychnine, and ephedrine),producing quaternary salts and other products(Phillipson and Bisset,1972).Similarly, the presence of traces of HCl may produce decomposition, dehydration, or isomerization in other compounds (Britton,et al.,1991).

Out of all the solvents, water is the most important of all extraction solvents (Mukherjee,2005).In our study fresh leaves juice showing more consistent antimicrobial activity.

It was clear that the methanol extract of selected medicinal plants exhibited high activity against the tested organisms rather than aqueous extract of those plants. Methanolic extracts of plants generally posses terpines and phenolics, which are reported by different workers as antimicrobial compounds ( Sanches, et al., 2005 ; Begum, et al., 2002 and Manach, et al., 2001). Acetone, Methanol and Petroleum ether extract of *Psidium guajava* showed pronounced activity against all the tested Gram positive and Gram negative microorganisms including *Pseudomonas* *aeruginosa*. (Mohamed etal.,1994; Kamath et al.,2008 and Dey et al.,2010).Guava leaf extract have been shown to be effective against many bacterial species known to cause diarrhea, including *S. aureus, E. coli* and other common enteropathogenic cultures (Jairaj et al.,1999; Coutino-Rodriguez et al., 2001). The methanolic extract of *P. guajava* (leaves) was the only agent showing significant inhibitory (and antidiarrheal) activities.

Guava is rich in tannins, phenols, triterpins, flavonoids, essential oils, saponins, carotenoids, lectins and all those compounds together showing antimicrobial activities (Kamath et al.,2008).Guava extracts of all polarity were found to be active against bacteria,indicating that more than one component may be responsible for the observed antimicrobial activity. Past research findings indicate the presence of polyphenolic compounds in guava, quercetin, avicularin and guaijaverin (Seshadri and Vasista, 1964) being the active antimicrobial components in guava leaf.

These findings support the traditional knowledge of local users about their selection of plant samples as antimicrobial agents and it is a preliminary scientific validation for the use of these plants for antibacterial activity. To promote proper conservation and sustainable use of such plant resources, awareness of local communities should be enhanced incorporating the traditional knowledge with scientific findings.The results of the present study also support the medicinal usage of the studied plants and suggest that some of the plant extracts possess compounds with antibacterial properties that can be used as antimicrobial agents in new drugs for the therapy of infectious diseases caused by pathogens. The most active extracts can be subjected to isolation of the therapeutic antimicrobials and undergo further pharmacological evaluation.

In conclusion Acetone, Methanol and Petroleum ether extracts of the plant parts showed antibacterial activity against disease-causing organisms and this suggest that constituents of the plants could be useful in chemotherapy. From the findings of this study, the following recommendations could be made; Firstly, there is a need to further isolate the active antibacterial agent (s) and secondly, it is necessary to determine toxicity of the active constituents, their side effects and pharmacokinetics effects.

**ACKNOWLEDGEMENT:**

We are thankful to University Grants Commission, New Delhi, India for financial assistance rendered to us.

**REFERENCES**

**Abdelrahim, S.I., Almagboul, A.Z., Omerb, M.E.A. and Elegamib,A.2002**.Antimicrobial activity of *Psidium guajava* L. *Fitoterapia.* ***73***: 713-715.

**Adriana,B., Almodovar, A. N. M., Pereiral, C.T. and Mariangela, T. A.2007.** Antimicrobial efficacy of *Curcuma zedoaria* extracts as assessed by linear regression compared with commercial mounthrinses. *Brazilion J Microbiol.***38:**440-445.

**Afolayan,A.J.2003.**Extracts from the shoots of Arctotis artotoides inhibit the growth of bacteria and fungi.*Pharm.Biol*.,**41:**22-25.

**Ahmad, I. and Beg, A.Z. 2001.** Antimicrobial and phytochemical studies on 45

*J.Ethnopharmacol.* ***74*:** 113-123.

**Al-Bari,M.A.,Sayeed,M.A.,Rahman,M.S. and Mossadik,** **M.A. 2006.** Characterization and antimicrobial activities of a phenolic acid derivative produced by Streptomyces bangladeshiensis,a novel species collected in Bangladesh.*Res.J Med &Medical Sci*.**1:**77-81.

**Bhat,S., Mercy Lobo., S., Chethan Kumar, K.V. and Sukeshand Chandrashekar, K.R. 2009.**Antimicrobial spectrum and phytochemical study of *Hopea parviflora* Beddome saw dust extracts. *Journal of Phytology*, **1(6):**469-474.

**Begum,S.,Hassan,S.I. and Siddiqui,B.S.2002.**Two new triterpenoids from the fresh leaves of *Psidium guajava .Phytochemistry*,**68:**1143-1152.

**Begum,S; Hassan,S.I.; Siddiqui, B.S.;Shaheen,F.;Ghayur,M.N.and Gilani,A.H.2002.** Triterpenoids from the leaves from *Psidium guajava. Phytochemistry*,**61,**399-403.

**Britton,G.Carotenoids,Dey,P.M.and Horborne, J.B.eds.** **1991.** *Methodsin Plant Biochemistry*, **Vol.7**, Academic,New York,474-517.

**Coutino-Rodriguez, R., Hernandez-Cruz, P. and Giles-Rios, H. 2001.** Lectins in fruits having gastrointestinal activity: their participation in the hemagglutinating property of *Escherichia coli* 0157:H7. *Arch Med Res*,**32:** 251-257.

**Dey,S.K.,Banerjee,D.,Chattapadhyay,S. and Karmarkar,K.B.2010.**Antimicrobial

activities of some medicinal plants of west Bengal.*Int.J.Pharma and Biosc.* **Vol-1(3):**1-10. **Dilhuydy,J.M.2003.**Patients attraction to complementary and alternative medicine (CAM): a reality which physicians can neither ignore nor deny.*Bulletin Cancer.* **90:**623-628.

**Doughari, J.H., El-mahmood, A.M. and Manzara, S. 2007.** Studies on the antibacterial activity of root extracts of *Carica papaya L*.,*African J. Microbiol. Res*. 037-041.

**Ellof, J.N. 1998.**Which extractant should be used for the screening and isolation of antimicrobial components from plants ? *J.Ethnopharmacol*.**60:**1-6.

**Erdogrul,O.T.,2002.**Antimicrobial activies of some plant extracts used in folklore medicine.*Pharmaceutical Biol*.**40:**269-273.

**Gnan, S.O. and Demello, M.T. 1999.** Inhibition of *Staphylococcus aureus* by aqueous *guajava* extracts. *J. Ethnopharmacol.* ***68:***103-108.

**Hassavi,D. and Kharma,A.2006.**Antimicrobial activity of medicinal plants against *Candida albicans*.*Journal Biological Sciences*,**6:**104-109

**Holetz, F.B., Pessini, G.L., Sanches, N.R., Cortez, D.A.G., Nakamur,C.V. and Dias Filho, B.P. 2002.** Screening of some plants used in the Brazilian folk medicine for the treatment of infectious diseases. *Mem Inst Oswaldo Cruz* ***97*:** 1027-1031.

**Jairaj, P., Khoohaswan, P., Wongkrajan, Y., Peungvicha, P., Suriywong, P., Sumal Saraya, M.L. and Ruangsomboon, O. 1999.** Anticough and antimicrobial activities of *Psidium guajava* Linn. Leaf extract. *J. Ethnopharmacol.* ***67*:** 203-212.

**Kamath, J.V., Rahul, N., Ashok Kumar, C.K. and Mohana Laksmi, S. 2008.***Psidium guajava* L:A review. *Int Jr Green Paharmacy*, **2(1):** 9-12.

**Kasamato, I.T., Nakabayasi,T. and Kida, H.,1995.** Screening of various plant extracts used in Ayurvedic medicine for inhibitory effects on human immunodeficiency virus type I(HIV-protease).*Phytotherpy Research*.**9:**180-184.

**Manach, C., Morand, C., Remesy, C. and Crespy, V. 2001.** Querctin 3-0 beta- glucoside is better absorbed that other querectin derivatives and is not present in rat plasma. Free Rad. Res. **33:** 667-676.

**Mohamed,S.,Hassan,Z. and Hamid,N. 1994.**Antimicrobial Activity of some Tropical Fruit Wastes.Pertanika *J.Trop.Agric.Sci*.**17(3):**219-227.

**Mukherjee,P.K. 2005.**Solvents used for extractions.*Quality control herbal drugsAn Approach to evaluation of Botanicals* Pub.Business Horizons. pp.390.

**Nascimento, G.G.F., Locatelli, J., Freitas, P.C.and Silva, G.L. 2000.**Antibacterial activity of plant extracts and phytochemicals on antibiotic-resistant bacteria. *Braz. J.Microbiol.* ***31*:** 247-256.

**Olajide,O.A.,Awe,S.O. and Makinde,J.M.1999.**Pharmacological studies on the leaf of *Psidium guajava.Fitoterapia*.**70:**25-31.

**Parekh, J. and Chanda, S. 2007.** In vitro antimicrobial activity and phytochemical analysis of some Indian medicinal plants. *Turk J. Biol*.**31:** 53-58.

**Parekh,J.,Darshana,J.and Sumitra,C.2005.**Efficacy of aqueous and methanol extracts of some medicinal plants for potential antibacterial activity.*Journal of Biology*.**29:**203-210.

**Parekh,J.,Karathia,N.and Chandra,S.2006.**Screening of some traditionally used medicinal plants for potential antibacterial activity.*Indian Journal of Pharmaceutical Sciences*.**68(6):**832-834.

**Perez, C., Paul, M.and Bazerque, P. 1990.** Antibiotic assay by agar well diffusion method. *Acta Bio. Med. Exp.* **15:** 113-115.

**Phillipson,J.D. and Bisset,N.G. 1972.** Quaternisation and oxidation of strychnine and brucine during plant extraction.*Phytochemistry*.**11:**2547-2553.

**Pretorius,C.J. and Watt, E. 2001.** Purification and identification of active components of *Carpobrotus edulis* L.*J. Ethnopharmacol*.**76:**87-91.

**Qadan, F., Thewaini, A.J., Ali, D.A., Afifi, R., Elkhawad, A. and Matalka ,K.Z. 2005.** The antimicrobial activities of *Psidium guajava* and *Juglans regia* leaf extracts to acne-developing organisms. *Am. J. Chin. Med.* ***33*:** 197-205.

**Sanches, N.R., Cortez, D.A.G., Schiavini, M.S,,Nakamura, C.V.and Dias Filho, B.P.** **2005.** An Evaluation of Antibacterial Activities of *Psidium guajava* (L.).*Braz. Arch.Biol.Tech. An Int. Jr.* **48(3):** 429-436.

**Shahidi Bonjar, G.H. 2004.**Evaluation of antimicrobial properties of Iranian medicinal plants against *Micrococcus luteus, Serratia marcescens, Klebsiella pneumoniae* and *Bordetell bronchiseptica.Asian J. Plant Sci.* **3:** 82-86, 2004.

**Seshadri, T.R. and K. Vasista. 1964.** Polyphenolic compounds of guava fruits. *Current Science* ***33:***334-335.

**Suresh,K., Deepa,P., Harisaranraj,R. and Achudhan,V. 2008.** Antimicrobial and phytochemical investigation of the leaves of *Carica papaya* L.,*Cynodon dactylon*(L.) *Pers.,Euphorbia hirta* L.,*Melia azedarach* L.and *Psidium guajava* L.: *Ethnobotanical Leaflets* **12:**1184-91.

**Santos,P.R.V., Oliveira, A.C.X., Tomassini,T.C.B. 1995.** Controle microbiogico de produtos fitoterapicos.*Rev.Farm.Bioquim*.**31:**35-38.

**Sofowara,A.1982.** Medicinal plants and antimicrobial activity .*Journal of Ethnopharmacology*.**100:**80-84.

**Vijaya, K.,Ananthan, S. 1997.** Microbiological screening of Indian medicinal plants with special reference to enteropathogens. *J. Altern Complement Med*. **3:**13-20.

**Voravuthikuchai, S., Lortheeranuwat, A., Jeeju, W., Sririrak, T., Phongpaichit, S.and Supawita, T. 2004.** Effective medicinal plants against entherohaemorragic *Escherichia coli* O157:H7. *J. Ethnopharmacol.* ***94*:** 49-54.