



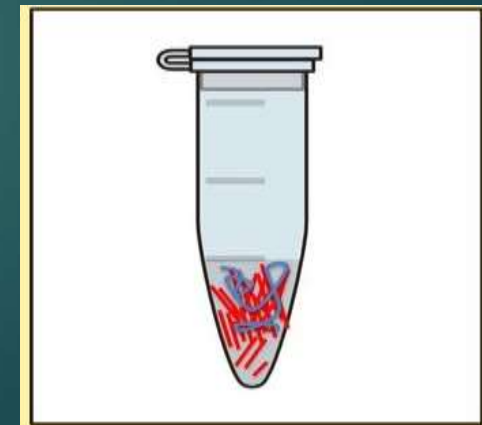
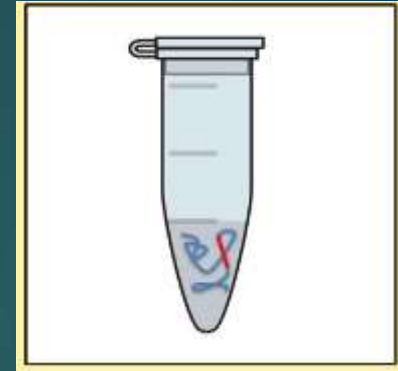
# DEPARTMENT OF BIOTECHNOLOGY

## PCR (Polymerase chain reaction)

Ms. Payal Talekar

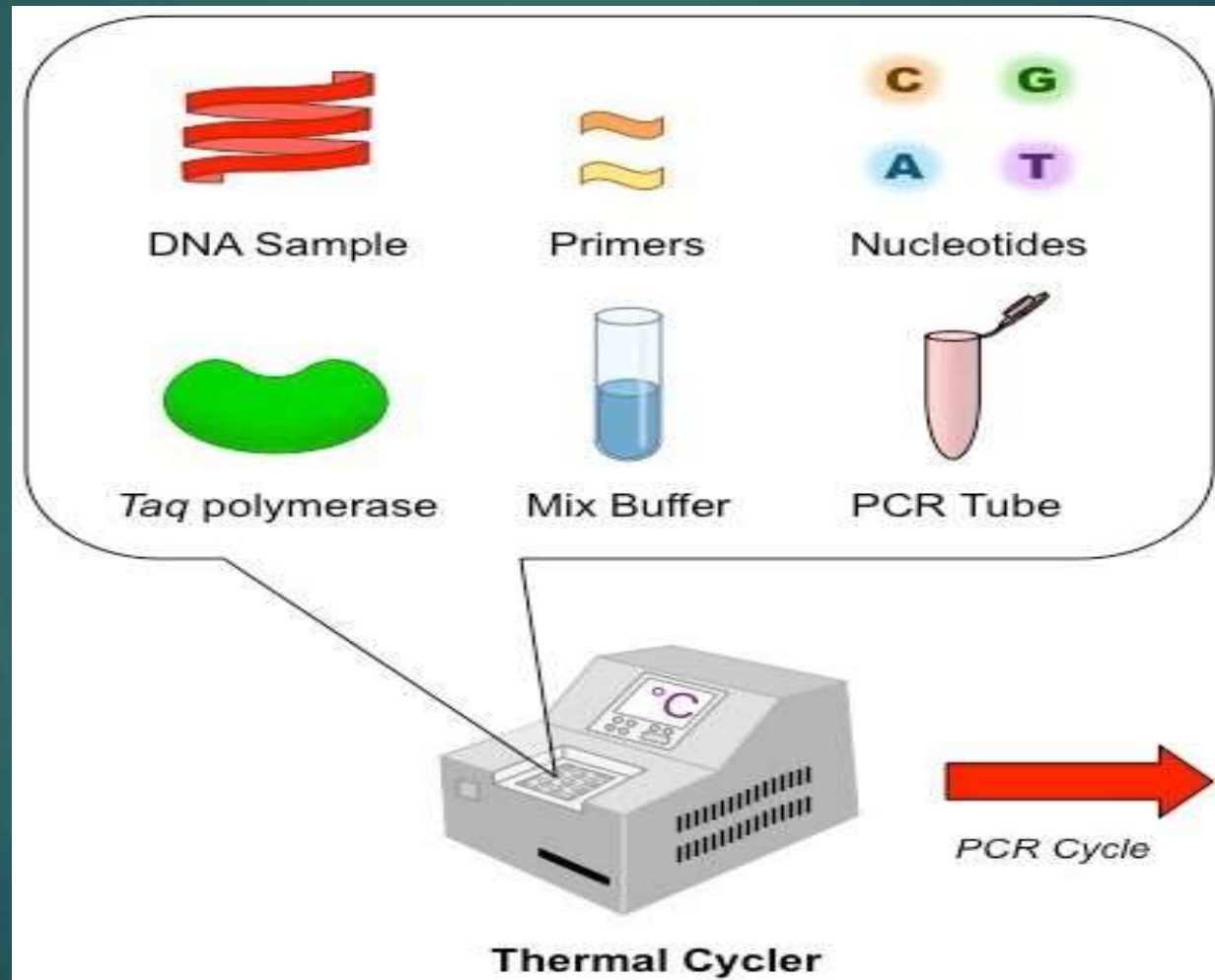
# PCR

- ❖ PCR is a means to amplify a particular piece of DNA .
- ❖ PCR can make billions of copies of a target sequence of DNA in short time.
- ❖ It is a laboratory level of DNA replication.





# Component of PCR



# PRIMER GUIDELINES

- Primer design
- Primer length
- GC %
- Melting temperature
- Annealing temperature.

# PRIMER GUIDELINES

## 1. Primer sequence:

- Must be complementary to flanking sequences of target region.
- Avoid:
  - Complementary sequences between primers.
  - Repeat (ex: ATATATAT) → misprime.
  - Runs (ex: AGCGGGGGAT) → misprime.
  - Mismatch at 3' end.

## 2. Primer length:

- It is generally accepted that the optimal length of primers is **18-25 bp**.
- Not too long nor too short

### 3. GC content:

- **GC%** = Number of G's and C's in the primer as a percentage of the total bases.
- Should be 40-60%.

### 4. GC clamp:

- Presence of G or C bases within the last five bases from the 3' end of primers.
- Not more than 2 G's or C's .

5'-CAACATAATAGCGACAACA**CTAGA**-3'

## 5. Melting temperature (T<sub>m</sub>):

- What is T<sub>m</sub>?
- Melting temperatures in the range of **50-60 °C** generally produce the best results.
- Maximum difference between primer pairs is 5°C.

The T<sub>m</sub> of the primer can be calculated by the following formula:

$$T_m = [(G + C) \times 4] + [(A + T) \times 2]$$

## 6. Annealing Temperature (T<sub>a</sub>):

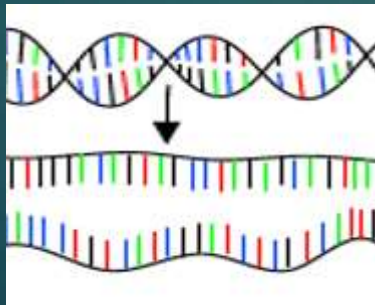
- The primer melting temperature is the estimate of the DNA-DNA hybrid stability and critical in **determining the annealing temperature**.
- Depends directly on **length** and **GC composition** of the primers.
- Too high T<sub>a</sub> → produce insufficient primer-template hybridization.
- Too low T<sub>a</sub> → lead to non-specific products caused by a high number of base pair mismatches.



# STEPS

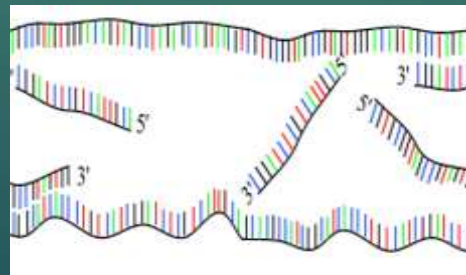
The double stranded DNA template DNA is denatured by heating, typically to 95 °C to separate the double stranded DNA

**Denaturation (92-95 °C)**



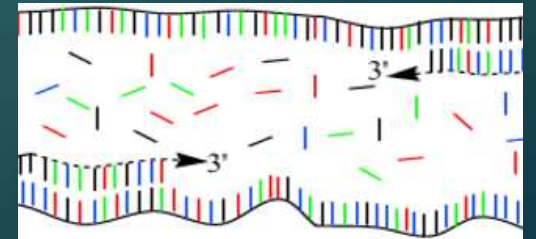
The reaction is rapidly cooled to an annealing temperature to allow the the oligonucleotide to hybridize the template.

**Annealing (50-65 °C)**



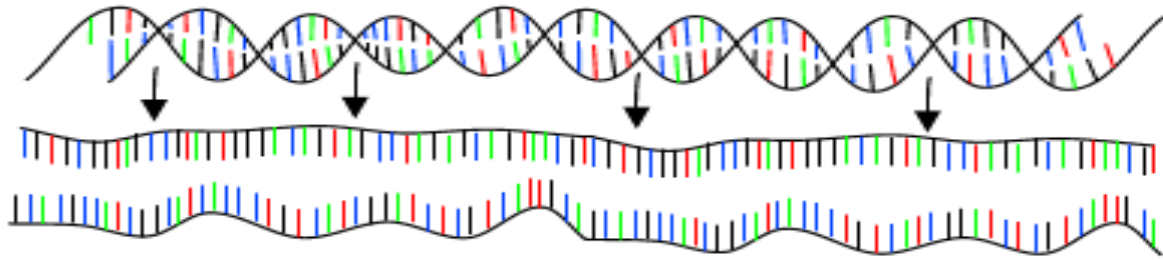
The reaction is heated to a temperature, typically 72 °C for efficient DNA synthesis by the thermostable DNA polymerase.

**Extension: (72 °C)**



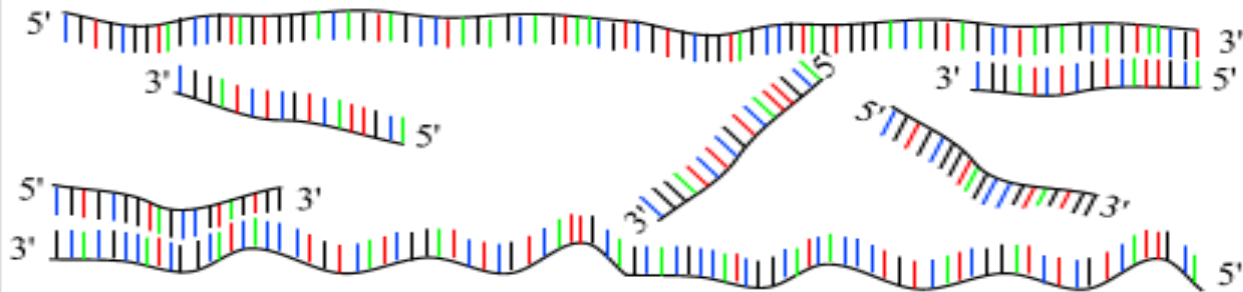
# PCR : Polymerase Chain Reaction

30 - 40 cycles of 3 steps :



**Step 1 : denaturation**

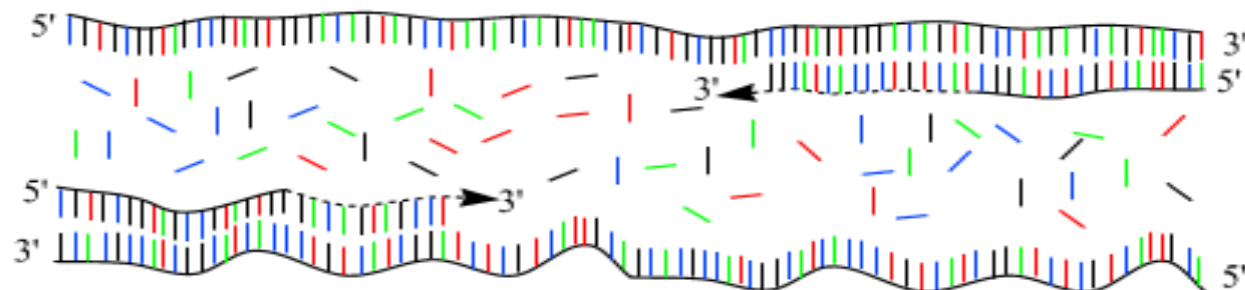
1 minut 94 °C



**Step 2 : annealing**

45 seconds 54 °C

**forward and reverse primers !!!**



**Step 3 : extension**

2 minutes 72 °C

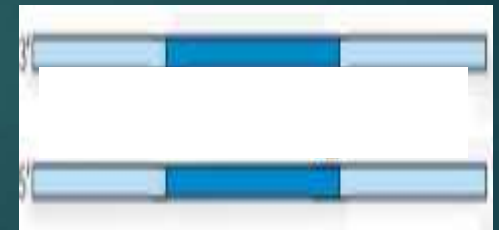
**only dNTP's**



# 1. Denaturation:

The double-stranded template DNA is denatured by heating, typically to **95°C**, to separate the double stranded DNA (why?).

Break the H bonds between the strands.



5' CATGCGATAAGAGTGATTGAGGT CCACCATGTTATCATGCGATAAGAGTGATTGAGGT CCACCATGTTATCATGCGATAAGAGTGATTGAGGT 3'

3' GTACGCTATTCTCACTAACTCCA GGTGGTACAATAGTACGCTATTCTCACTAACTCCA GGTGGTACAATAGTACGCTATTCTCACTAACTCCA 5'



## 2. ANNEALING:

- The reaction is rapidly cooled to the primer annealing temperature (50-65 °C) to allow the oligonucleotide primers to hybridize to single stranded template.
- Primer will anneal only to sequences that are complementary to them (target sequence).



# Annealing:

5' CATGCGATAAGAGTGATTGAGGT CCACCATGTTATCATGCGATAAGAGTGATTGAGGT CCACCATGTTATCATGCGATAAGAGTGATTGAGGT 3'  
3' GGTGGTACAATAGTACGCTATT 5'

CCACCATGTTATCATGCGA 3'  
3' GTACGCTATTCTCACTAACTCCA GGTGGTACAATAGTACGCTATTCTCACTAACTCCA GGTGGTACAATAGTACGCTATTCTCACTAACTCCA 5'

Forward primer: 5' CCACCATGTTATCATGCGA 3'

Reverse primer: 3' GGTGGTACAATAGTACGCTATT 5'

# EXTENSION:

- The reaction is heated to a temperature depends on the DNA polymerase used.
- **Commonly** a temperature of 72°C is used with this enzyme.
- This means that 72°C is the optimum temperature of DNA polymerase.
- At this step the DNA polymerase synthesizes a new DNA strand complementary to the DNA template.

5' CATGCGATAAGAGTGATTGAGGT CCACCATGTTATCATGCGATAAGAGTGATTGAGGT CCACCATGTTATCATGCGATAAGAGTGATTGAGGT 3'  
← TATTCTCACTAACTCCA GGTGGTACAATAGTACGCTATT 5'

5' CCACCATGTTATCATGCGA TAAGAGTGATTGAGGT →  
3' GTACGCTATTCTCACTAACTCCA GGTGGTACAATAGTACGCTATTCTCACTAACTCCA GGTGGTACAATAGTACGCTATTCTCACTAACTCCA 5'

Taq DNA polymerase



# 3. Extension :

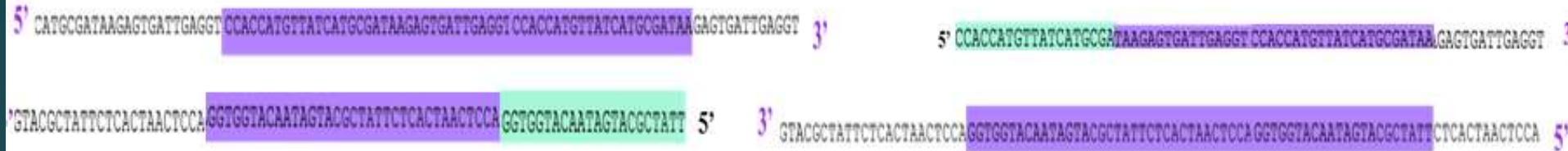


Cycle # 1:  
1 DNA amplified to 2 DNA

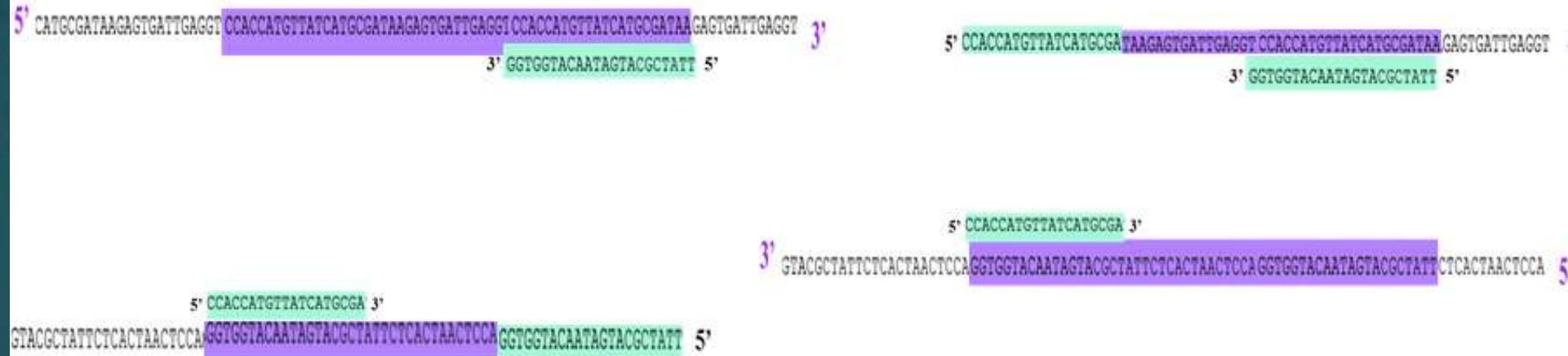


# Cycle 2

## 1. Denaturation

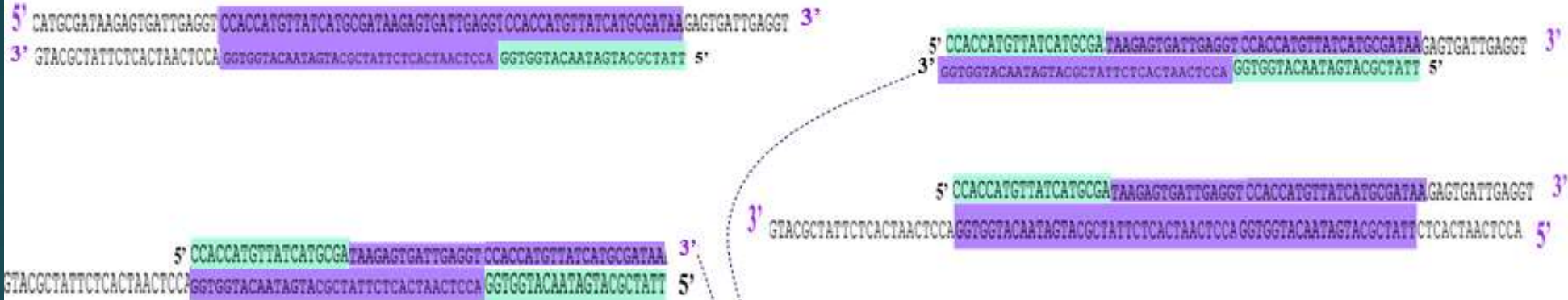


## 2. Annealing



# Cycle 2

## 3. Extension



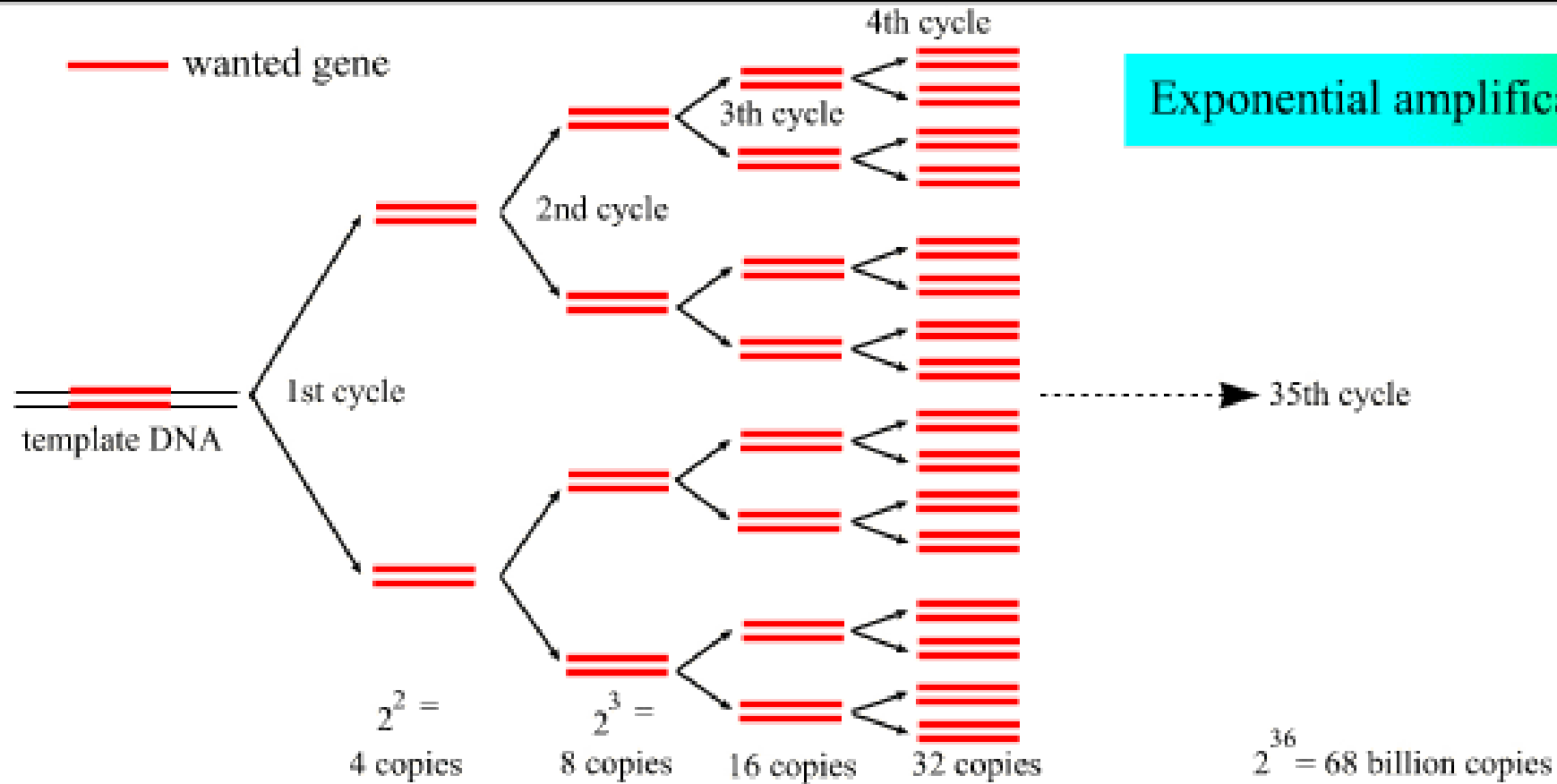
# Cycle 3



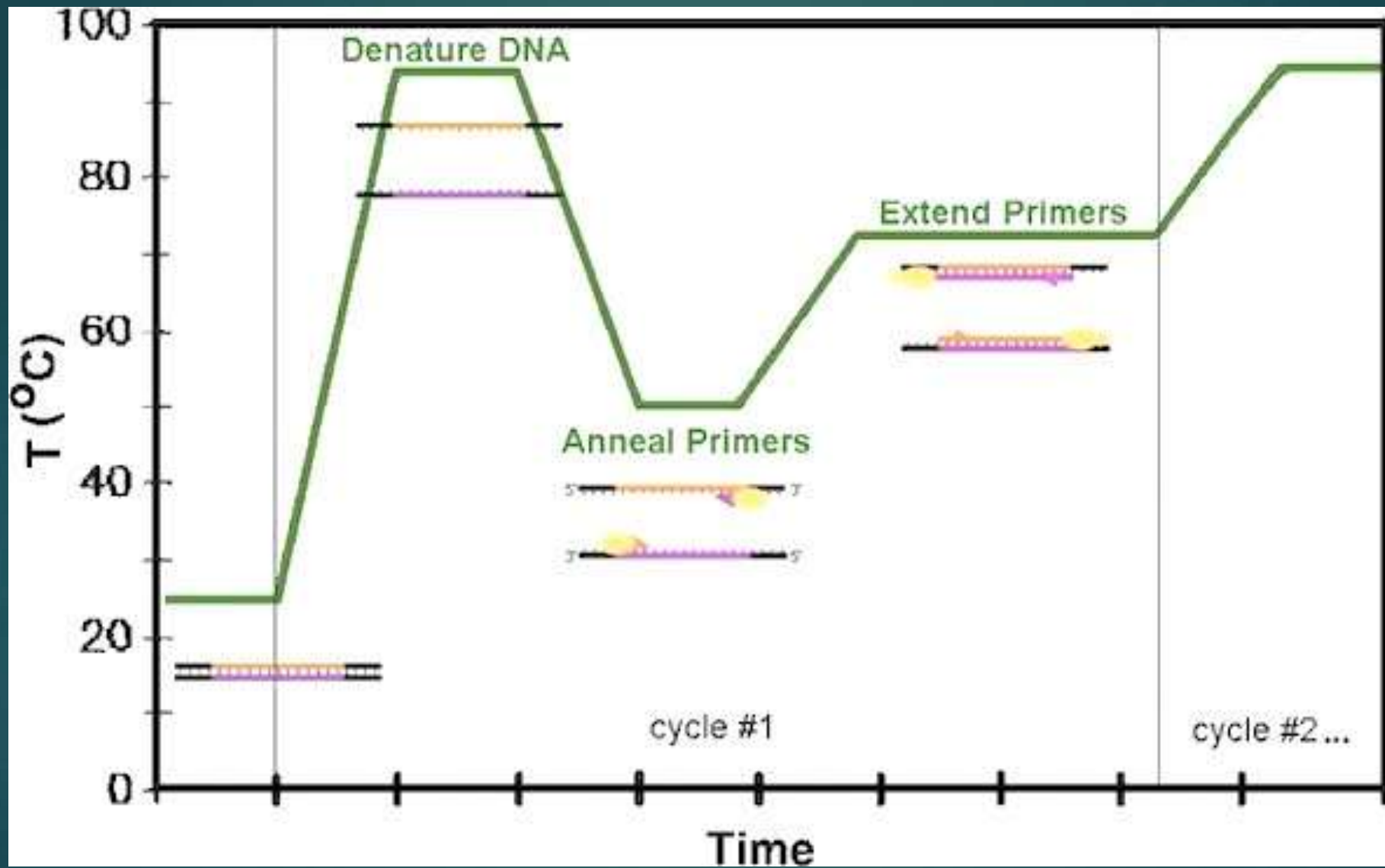
**Target sequence**  
Appeared after three cycles and  
start to accumulate



**After 30 cycles:**  
 $2^{30}$  copy of target DNA !!



(Andy Vierstraete 1999)



# Applications of PCR

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graph TD; A[Applications of PCR] --> B[Basic Research]; A --> C[Applied Research];
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## Basic Research

- Mutation screening
- Drug discovery
- Classification of organisms
- Genotyping
- Molecular Archaeology
- Molecular Epidemiology
- Molecular Ecology
- Bioinformatics
- Genomic cloning
- Site-directed mutagenesis
- Gene expression studies

## Applied Research

- Genetic matching
- Detection of pathogens
- Pre-natal diagnosis
- DNA fingerprinting
- Gene therapy