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EVALUATION ON ANTIMICROBIAL ACTIVITIES OF SELECTED INDIAN MEDICINAL PLANTS

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ABSTRACT:

The four plants were selected for this research namely *Cassia* auriculata, *Glycyrrhiza glabra*, *Piper longum and Syzygium* aromaticum based on ethno botanical information, literature, references and therapeutic significance. These weremostly reported for antidiabetic, anti- cancer, anti-inflammatory activities and for several other therapeutic indications. Though several researchers also reported antimicrobial activities for these plants, most of these plants have not been fully studied for their antimicrobial potential and exploited as antimicrobials, especially as drug potentiators, or resistance modifying agents, or synergistic agents. In this article, antimicrobial activities of selected Indian medicinal plants has been evaluated. **KEYWORDS:**Antimicrobial, Indian, Medicinal, Plants.

INTRODUCTION:

One of the main therapeutic challenges faced by antibiotic therapy is the emergence of multidrug resistance, particularly in bacteria. [1] To combat the diseases caused by these microorganisms, researchers have been compelled to discover and develop new medications. [2] However, the past few years have witnessed a decrease in the development of new antibiotics due to the time-consuming, expensive, and labor-intensive nature of isolating the new antimicrobial agents with unique modes of action from microbes. [3] Furthermore, the limited shelf life of newly introduced antibiotics stems from their frequent updates every two to three years. [4] Many of the antibacterial agents that are now in use have negative side effects that include toxicity, immunosuppression, hypersensitivity, and tissue residue that pose a risk to public health. [5] Moreover, the more recent broad spectrum antibiotics are more expensive, making them inaccessible to the impoverished. [6] These drawbacks make it difficult to use the antibacterial drugs that are now on the market therapeutically, which makes it necessary to look for alternate treatments for illnesses caused by different bacteria. [7,8] As the world shifts towards non-toxic and environmentally friendly products, we should prioritize the creation of pharmaceuticals from traditional medicinal plants for the treatment of many human and animal diseases. [9, 10]

EXPERIMENTAL METHODOLOGY:

The experimental methodology based on the following protocol:

- Selection of Plants
- Identification of Plant Parts
- Plant Extracts Preparation

- \geq Yield Percentage of Solvent Extracts
- ➢ Test Organism:

The following microorganisms was used for antimicrobial activity of the plant extracts.

- Gram-positive strains (ATCC):
- Bacillus subtilis
- Staphylococcus aureus
- Micrococcus luteus •

Gram-negative strains (ATCC):

- Escherichia coli
- Pseudomonas aeruginosa .
- Salmonella typhi
- Culture Media Composition of Different Media: \triangleright

The culture media used for anti-microbial susceptibility testing of the bacterial & fungal strains in the present study were as follows:

- Nutrient Broth
- Nutrient Agar
- Potato Dextrose Broth
- Potato Dextrose Agar
- Preparation of Inoculum Growth Media \geqslant
- \triangleright Preparation of Turbidity standard
- Preparation of dried filter paper discs
- Preparation of dried filter paper discs
- Antibiotic Discs
- AAA Antibiotic Susceptibility Test
- Evaluation of Anti-Bacterial Activity of Plant Extracts
- \triangleright Minimum Inhibitory Concentration
- \triangleright Minimum Bactericidal Concentrations

RESULTS:

Cassia auriculata

vitro antibacterial (activity) of four therapeutic In potential plants selected randomly assessed (medicinal plants) leaves extracts have been by the method of agar well diffusion assay (AWDA).

	Zone of Inhibition (mm)										
Extracts	B. s.	S. a.	M. l.	E.c	P.a	S.t.					
Methanol	20	12	18	17	13	16					
Ethanol	17	12	16	15	11	14					
Aqueous	12	09	13	16	08	12					
reptomycin	23	22	23	25	16	20					
DMSO	00	00	00	00	00	00					

Table 1: Antibacterial activity of *Cassia auriculata* plant extracts

Streptomycin (+ ve control) DMSO (- ve control)



Figure 1: Antibacterial activity of Cassia auriculata plant extracts

Glycyrrhiza glabra

Table 2: Antibacterial activity of Glycyrrhiza glabra stem bark extracts

				Zon	e of Inhibit	tion (mm)
Extracts	B. s.	S. a.	M. l.	E.c	P.a	S.t.
Methanol	21	25	18	13	00	00
Ethanol	19	22	16	00	00	00
Aqueous	17	20	14	00	00	00
treptomycin	24	27	22	23	16	17
DMSO	00	00	00	00	00	00



Figure 2: Antibacterial activity of *Glycyrrhiza glabra* stem bark extracts

				Zon	e of Inhibit	tion (mm)
Extracts	B. s.	S. a.	M.I	E.c	P.a	S.t.
Methanol	14	12	00	13	00	00
Ethanol	12	10	00	09	00	00
Aqueous	00	00	00	00	00	00
treptomycin	21	23	20	22	16	17
DMSO	00	00	00	00	00	00

Piper longum



Figure 3:Antibacterial activity of *Piper longum* stem dried root extracts

Syzygium aromaticum

 Table 4:Antibacterial activity of Syzygium aromaticum
 dried flower extracts

bition (mm)						
Extracts	B. s.	S.a	M. l	E.c	P.a	S.t.
Methanol	19	21	22	20	13	17
Ethanol	16	17	19	17	18	12
Aqueous	15	14	17	15	08	13
Streptomycin	21	20	23	22	16	19
DMSO	00	00	00	00	00	00



Figure 4: Antibacterial activity of Syzygium aromaticumdried flower extracts

					MI	C (mgml-1)
Extracts	B. s.	S.a	M. l	E.c	P.a	S.t.
Methanol	25	50	-	50	-	-
Ethanol	50	100	-	50	-	-
Aqueous	-	-	-	-	-	-

Table 5: MIC (mgml⁻¹) of *Cassia auriculata* plant extracts



Figure 5:MIC (mgml⁻¹) of Cassia auriculata plant extracts

					MIC	(mgml-1)
Extracts	B. s.	S.a	M. l	E.c	P.a	S.t.
Methanol	25	12.5	12.5	12.5	-	-
Ethanol	50	50	12.5	12.5	-	-
Aqueous	100	100	25	-	-	-

 Table 6: MIC (mgml⁻¹) of Glycyrrhiza glabra stem bark extracts



Figure 6: MIC (mgml-1) of Glycyrrhiza glabra stem bark extracts

					MI	C (mgml ⁻¹)
Extracts	B. s.	S.a	M. l	E.c	P.a	S.t.
Methanol	12.5	12.5	25	12.5	50	25
Ethanol	25	25	25	12.5	50	50
Aqueous	25	25	50	12.5	100	100

Table 7: MIC	$(mgml^{-1})$	of Piper	longum	root extracts
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Figure 7: MIC (mgml⁻¹) of *Piper longum* root extracts **Table 8: MIC (mgml⁻¹) of** *Syzygium aromaticum* flower extracts

					MI	C (mgml ⁻¹)
Extracts	B. s.	S.a	M. l	E.c	P.a	S.t.
Methanol	25	25	100	50	50	25
Ethanol	50	100	50	50	100	100
Aqueous	50	50	-	100	-	-



Figure 8: MIC (mgml⁻¹) of *Syzygium aromaticum* flower extracts

					MB	C (mgml- ¹)
Extracts	B. s.	S.a	M. l	E.c	P.a	S.t.
Methanol	50	-	100	50	-	-
Ethanol	100	-	100	100	-	-
Aqueous	-	-	-	-	-	-

Table 9: MBC (mgml-¹) of *Cassia auriculata* plant extracts



Figure 9: MBC (mgml⁻¹) of *Cassia auriculata* plant extracts

Table 10: MBC (mgml ⁻¹) of <i>Glycyrrhiza glabra</i> stem bark extracts									
MBC (mgml- ¹)									
Extracts	B. s.	S.a	M. l	E.c	P.a	S.t.			
Methanol	25	25	12.5	25	50	25			
Ethanol	50	25	25	25	100	50			
Aqueous	50	50	25	50	25	50			





Figure 10: MBC (mgml⁻¹) of *Glycyrrhiza glabra* stem bark extracts

MBC (mgml- ¹)							
Extracts	B. s.	S.a	M. l	E.c	P.a	S.t.	
Methanol	25	50	50	100	50	100	
Ethanol	50	100	100	50	50	100	
Aqueous	100	-	-	-	-	-	

Table 11: MBC (mgml⁻¹) of *Piper longum* root extracts



Figure 11: MBC (mgml⁻¹) of *Piper longum* root extracts

MBC (mgml ⁻¹)							
Extracts	B. s.	S.a	M. l	E.c	P.a	S.t.	
Methanol	25	50	50	25	50	50	
Ethanol	50	50	100	50	100	100	
Aqueous	50	100	100	50	100	100	

Table 12: MBC (mgml ⁻¹) of Syzygr	ium aromaticum flower extracts
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Figure 12: MBC (mgml⁻¹) of *Syzygium aromaticum* flower extracts

CONCLUSION:

From the above results, it was found that the four medicinal plants namely *Cassia auriculata*, *Glycyrrhiza glabra*, *Piper longum and Syzygium aromaticum* have potent antibacterial as well as antimicrobial activities.

REFERENCES:

- 1. Karahan, F. et al. (2016) Antimicrobial and antioxidant activities of medicinal plant *Glycyrrhiza glabra var. glandulifera* from different habitats.*Biotechnology & Biotechnological Equipment*, 30(4), 797-804.
- 2. Syed, S.R., Shekshavali, T., Ahamed, S.S. (2018). A Review on *Cassia auriculata. Res. J. Pharmacology and Pharmacodynamics*, 10(3), 141-145.
- 3. Batiha, G.E.S. et al. (2020). *Syzygium aromaticum* L. (Myrtaceae): Traditional Uses, Bioactive Chemical Constituents, Pharmacological and Toxicological Activities.*Biomolecules*, 10(2), 202.
- 4. Seema Yadav et al. (2020). Syzygium Aromaticum (Clove): A Review on Various Phytochemicals and Pharmacological Activities in Medicinal Plant.*World Journal of Pharmaceutical Research*, 9 (11), 349.363.
- 5. Meena, V. et al. (2019). Cassia auriculata: A healing herb for all remedy. *Journal of Pharmacognosy and Phytochemistry*, 8(3), 4093-4097.
- 6. Ashalatha, M & Sannappanawar, R.B. (2015). A Review Article on Pippali (*Piper longum* Linn), *International Ayurvedic Medical Journal*, 3(9), 2841-2849.
- 7. Pandey, D.K. (2018). Piper longum: A concise review on Botany, Phytochemistry and Pharmacology. *Journal of Emerging Technologies and Innovative Research*, 5(12), 711-717.
- 8. Pastorino, G. et al. (2018). Liquorice (Glycyrrhiza glabra): A phytochemical and pharmacological review.*Phytother Res*, 32(12), 2323–2339.
- 9. Chauhan, N., Uniyal, P., Chauhan, R., Singh, C., Kumar, D. (2019). In Vitro Antibacterial Effects of Piper longum Fruit Extracts on Human Pathogens and Phytochemical Analysis. *International Journal of Research and Analytical Reviews*, 6 (1), 282-288.
- 10. Saritha, P.&Devi, U. A. (2017). Medicinal Properties of Telangana State Flower Tangedu (Cassia Auriculata Linn). *World Journal of Pharmaceutical Research*, 6(8), 1597-1605.